

# **Syllabus of BIOINF 528 (2017 Fall, Bioinformatics)**

## **Course Name:**

Advanced Applications of Bioinformatics

## **Course Description:**

This course introduces fundamental concepts and methods for bioinformatics and the advanced applications. Topics covered include bioinformatics database, sequence and structure alignments, protein folding and protein structure prediction, protein-protein interaction, Monte Carlo simulation, and structural determination of macromolecules (X-ray crystallography and NMR). Emphasis is on the understanding of the concepts taught and their practical utilization, with the objective of helping student use cutting-edge bioinformatics tools/methods to solve problems in their own research.

## **Instructor:**

Yang Zhang, Email: [zhng@umich.edu](mailto:zhng@umich.edu), Phone: 734-647-1549

## **Invited lecturers:**

Philip Andrews, Tomek Cierpicki, John Tesmer

## **Schedule and location:**

9:00 am - 12:00 noon, Friday; #2036 Palmer Commons

## **Homework, lab & Exams:**

There will be homework assignments, including code writing and literature reading. Homework and lab work constitutes 60% of your grade; the final exam constitutes another 40%.

## **Student evaluation and grades:**

A = 90-100%      B = 80-89      C = 70-79      D = 60-69      F = below 60

## **Textbook:**

No textbook is required for this course. Assigned materials will be either handed out to the class or posted on the blackboard website.

# Table of content

## **PART I: BIOINFORMATICS BASICS**

### **1. Bioinformatics databases**

- 1.1. Introduction
  - 1.1.1. Motivation
  - 1.1.2. Type of databases
- 1.2. Nucleotide sequence databases
  - 1.2.1. EMBL
  - 1.2.2. GeneBank
  - 1.2.3. DDBJ
- 1.3. Protein sequence databases
  - 1.3.1. How protein sequences are determined
    - 1.3.1.1. DNA/mRNA coding
    - 1.3.1.2. Edman degradation reaction
    - 1.3.1.3. Mass spectrometry
  - 1.3.2. SwissProt/TrEMBL
  - 1.3.3. PIR
  - 1.3.4. UniProt
- 1.4. Protein structure databases
  - 1.4.1. Protein Data Bank
  - 1.4.2. SCOP
  - 1.4.3. CATH
- 1.5. Other relevant databases
  - 1.5.1. Pfam
  - 1.5.2. KEGG
  - 1.5.3. PROSITE
  - 1.5.4. BioLiP

### **2. Pair-wise sequence alignments and database search**

- 2.1. Biological motivation
- 2.2. Pairwise alignments
  - 2.2.1. Scoring matrix
    - 2.2.1.1. PAM
    - 2.2.1.2. BLOSUM
  - 2.2.2. Gap penalty
- 2.3. Dynamic programming
  - 2.3.1. Needleman-Wunsch
  - 2.3.2. Smith-Waterman
- 2.4. Heuristic methods
  - 2.4.1. FASTA
  - 2.4.2. BLAST
- 2.5. Statistics of sequence alignment score
  - 2.5.1. E-Value
  - 2.5.2. P-Value

### **3. Phylogenetic tree & multiple sequence alignments**

- 3.1. Neighbor-joining method and phylogenetic tree
- 3.2. How to construct multiple sequence alignment?
  - 3.2.1. ClustalW
  - 3.2.2. PSI-BLAST
  - 3.2.3. Hidden Markov Models
    - 3.2.3.1. Viterbi algorithm
    - 3.2.3.2. HMM based multiple-sequence alignment
      - 3.2.3.2.1. HMMER
      - 3.2.3.2.2. SAM
- 3.3. Sequence profile & profile based alignment
  - 3.3.1. Profile-to-sequence alignment
  - 3.3.2. Profile-to-profile alignment
- 4. **Protein structure alignments**
  - 4.1. What is sequence-dependent structure superposition?
    - 4.1.1. RMSD
    - 4.1.2. TM-score
  - 4.2. What is sequence-independent structure alignment?
  - 4.3. Different structure alignment algorithms
    - 4.3.1. DALI
    - 4.3.2. CE
    - 4.3.3. TM-align
  - 4.4. How to define the fold of proteins?
  - 4.5. Number of protein folds in PDB

## **PART II: PROTEIN FOLDING AND PROTEIN-PROTEIN INTERACTIONS**

- 5. **Protein secondary structure predictions**
  - 5.1. What is protein secondary structure?
  - 5.2. Hydrogen bond
  - 5.3. How to define a secondary structure element?
  - 5.4. Methods for predicting secondary structure
    - 5.4.1. Chou and Fasman method
    - 5.4.2. PHD
    - 5.4.3. PSIPRED
    - 5.4.4. PSSpred
- 6. **Monte Carlo Simulation**
  - 6.1. Introduction: why Monte Carlo simulation?
  - 6.2. Monte Carlo Sampling of Probabilities
    - 6.2.1. Random number generator
      - 6.2.1.1. How to test a random number generator?
    - 6.2.2. Sampling of rectangular distributions
    - 6.2.3. Sampling of probability distribution
      - 6.2.3.1. Reverse transform method
      - 6.2.3.2. Rejection sampling method
  - 6.3. Boltzmann distribution
  - 6.4. Metropolis method

- 6.5. Advanced Metropolis methods
  - 6.5.1. Replica exchange simulation
  - 6.5.2. Simulated annealing
- 7. Protein tertiary structure modeling**
  - 7.1. Basic concepts
  - 7.2. Ab initio modeling
    - 7.2.1. Anfinsen thermodynamic hypothesis
    - 7.2.2. Molecular dynamics simulation
      - 7.2.2.1.CHARMM
      - 7.2.2.2.AMBER
    - 7.2.3. Knowledge-based free modeling
      - 7.2.3.1.Bowie-Eisenberg approach
      - 7.2.3.2.ROSETTA
      - 7.2.3.3.QUARK
      - 7.2.3.4.Why is beta-protein so difficult to fold?
  - 7.3. Comparative modeling (homology modeling)
    - 7.3.1. Principle of homology modeling
    - 7.3.2. Modeller
  - 7.4. Threading and fold-recognition
    - 7.4.1. What is threading?
    - 7.4.2. Threading programs
      - 7.4.2.1.Bowie-Luthy-Eisenberg
      - 7.4.2.2.HHpred
      - 7.4.2.3.MUSTER
    - 7.4.3. Meta-server threading
      - 7.4.3.1.3D-jury
      - 7.4.3.2.LOMETS
  - 7.5. Combined modeling approaches
    - 7.5.1. TASSER/I-TASSER
  - 7.6. CASP: A blind test on protein structure predictions
- 8. Principle of X-ray Crystallography & Molecular Replacement**
  - 8.1. What is X-ray Crystallography
  - 8.2. Why can a wave be represented by  $\exp(i\alpha)$  ?
  - 8.3. How to calculate scattering on two electrons?
  - 8.4. What is Laue condition?
  - 8.5. What is Bragg's law?
  - 8.6. How to calculate electron density of crystal?
  - 8.7. What is Patterson function?
  - 8.8. How to calculate electron density of crystal?
  - 8.9. What is the idea of Molecular Replacement?
  - 8.10. How to judge quality of MR?
  - 8.11. What are often-used software for MR?
- 9. Introduction to nuclear magnetic resonance (NMR)**
  - 9.1. Basic magnetic property of nuclei
    - 9.1.1. Magnetic moment
    - 9.1.2. Nuclei in external magnetic field

- 9.1.3. Nuclear shielding of magnetic field
- 9.2. Chemical shift
- 9.3. NMR spectrum
  - 9.3.1. Correlation spectroscopy (COSY)
  - 9.3.2. Heteronuclear single-quantum correlation spectroscopy (HSQC)
  - 9.3.3. Nuclear Overhauser effect spectroscopy (NOESY)
- 9.4. From NOE to 3D structure model
- 10. Protein function and structure-based function annotation**
  - 10.1. Gene ontology
  - 10.2. Enzyme classification
  - 10.3. Ligand-protein interaction
  - 10.4. Structure-based function annotation
    - 10.4.1. FindSite
    - 10.4.2. COFACTOR
    - 10.4.3. COACH
- 11. Modeling of protein-protein interactions**
  - 11.1. Experimental identification of protein-protein interactions
    - 11.1.1. Yeast two-hybrid assay
    - 11.1.2. High-throughput mass spectrometry
    - 11.1.3. Interaction networks and system biology
  - 11.2. Protein-protein quaternary structure modeling
    - 11.2.1. Basic concepts
      - 11.2.1.1. Degrees of freedom
      - 11.2.1.2. Presentation of protein conformations
      - 11.2.1.3. Hydrophobicity factor
      - 11.2.1.4. Shape complementary
      - 11.2.1.5. Docking Scoring function
    - 11.2.2. Protein-protein docking algorithms
      - 11.2.2.1. Fast Fourier Transformation (FFT)
      - 11.2.2.2. Semi-flexible docking: Side-chain refinement
      - 11.2.2.3. Clustering and refinement
    - 11.2.3. Homology modeling
    - 11.2.4. Multiple-chain threading
      - 11.2.4.1. COTH
      - 11.2.4.2. SPRING
    - 11.2.5. Monte Carlo docking simulation
    - 11.2.6. CAPRI: A blind test on protein-protein docking

**PART III: Selected Topics (by invited speakers)**

**Prof. Philip Andrews:**

Introduction to mass spectrometry and proteomics

**Prof. Tomek Cierpicki:**

NMR spectroscopy for protein structure determination

**Prof. Daniel Southworth:**

Cryo-electron microscopy for protein structure determination

**Prof. Matthew Young:**

X-ray crystallography for protein structure determination